

A photograph of a sliced apple on a wooden cutting board. The apple is cut in half, revealing its core and seeds. Several capsules are placed inside the core of the apple. The background is a wooden surface with a vertical grain.

African Journal of Pharmacy and Pharmacology

Volume 10 Number 40, 29 October, 2016
ISSN 1996-0816



*Academic
Journals*

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Full Length Research Paper

Analgesic and central nervous system depressant activities of methanol extract of *Ziziphus rugosa* Lam. Leaves

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Received 6 August, 2015; Accepted 3 September, 2015

The present investigation was designed to evaluate the analgesic and central nervous system (CNS) depressant activities of the methanolic extract of *Ziziphus rugosa* Lam. in rat model. The analgesic activity of the extract was examined using acetic acid-induced writhing test (chemically induced pain) at the doses of 200 and 300 mg/kg body weight. Further, the CNS depressant activity of the plant extract was evaluated by open field and hole cross tests at the doses of 300 and 500 mg/kg body weight. In the analgesic activity test, the methanolic extract of *Z. rugosa* showed significant ($p < 0.01$) antinociceptive activity in a dose dependent manner. The extract at the dose of 300 mg/kg exhibited 51.87% inhibition of writhing response which was comparable to the reference drug Indomethacin (55.20%). On the other hand, the plant extract also demonstrated significant ($p < 0.01$) dose-dependent reduction of locomotor and exploratory activities in the open field and hole cross tests. This analgesic and CNS-depressant activity of the extract might be due to the presence of biologically important chemical compounds.

Key words: Central nervous system (CNS)-depressant, antinociceptive, hole cross tests, writhing, locomotor activity.

INTRODUCTION

Pain is the part of a defensive response against dysfunction of an organ or imbalance in its functions towards potentially dangerous stimulus. Pain management has become the focus of global scientific research because of its inference in virtually all human and animal diseases (Sarker et al., 2012). Many drugs are used to relieve the pain and most of them produce some side-effects on the physiology of the body (Devraj and Karpagam, 2011). As a result, searching of new

analgesic drugs from traditionally used plant species as pain killers should still be a rational strategy. The plant *Ziziphus rugosa* Lam. is a large straggling armed shrub with large elliptic usually subcordate leaves and belongs to the family Rhamnaceae. The flowers are paniculated and fruits are small drupe, glabrous and become white when ripe. Traditionally bark is used as astringent and antidiarrhoeal while flowers are used for menorrhagia and hemorrhage. Stem and fruit are hypotensive. Fruit is used

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in the treatment of rheumatism and hypotension. The root bark of the plant have been reported to exhibit analgesic, anti-inflammatory, antibacterial and antifungal activities (Abhimany and Pratiksha, 2010; Rambabu et al., 2011). The seed extract has been reported for anxiolytic and cytotoxic properties (Prashith et al., 2011; Gawande et al., 2011). Cytotoxicity, antimicrobial and antioxidant properties (Sazzad et al., 2013) have also been reported from leaves. The fruit pericarp is known to have antibacterial, insecticidal and free radical scavenging properties (Kekuda et al., 2011). Rugosanine-A, a cyclopeptide alkaloid (Pandey et al., 1988) and three flavonoids - kaempferol-4'-methylether, luteolin and luteolin-7-O-glucoside (Singh et al., 2009) have been isolated from the stem bark of the plant.

A new glycoside zizyphoside has been isolated along with betulic, oleanolic, aliphatic and 2- α -hydroxyursolic acids; zizyphoside on hydrolysis yielded altered aglycone, ebelin lactone (Bakshi et al., 1999). Due to the absence of report of the *Ziziphus rugosa* leaves on central nervous system (CNS) activity, an effort has been made as part of our regular research program (Rahman et al., 2008; Ferdous et al., 2012; Khan et al., 2015) to scientifically evaluate the CNS depressant and analgesic effects of methanolic extract of *Z. rugosa* leaves in model rat.

MATERIALS AND METHODS

Plant

The leaves of plant *Z. rugosa* were collected from Gazipur in the month of March, 2011 and identified in the Bangladesh National Herbarium, Dhaka, where a voucher specimen (No. DACB-34479) has been mentioned for future reference.

Preparation of extract

The leaves were first washed with running water to remove adhering dirt, then dried in an electric oven at 45°C and powdered with a mechanical grinder, passing through sieve #40, and stored in a tight container. The powdered material (1 kg) was taken in a clean, flat bottomed glass container and soaked in methanol for seven days. The whole mixture was then filtered through a piece of clean, white cotton bed followed by Whatman filter paper number 1. The total filtrate was concentrated, *in vacuo* at 40°C to render brownish red colored mass (390 g).

Drugs and chemicals

The active drugs Indomethacin and Diazepam were obtained from Square Pharmaceuticals Ltd., Bangladesh. Acetic acid was obtained from Merck, Germany. Tween-80 was procured from BDH Chemicals, UK. Normal saline solution was purchased from Beximco Infusion Ltd., Bangladesh. All chemicals used were of analytical reagent grade.

Animals

Long-Evan's rats of either sex weighing about 120 to 170 g were

used for the experiment. The rats were purchased from the Animal Resources Branch of the International Centre for Diarrhoeal Disease and Research, Bangladesh (ICDDR, B). They were kept in standard environmental condition (at 24.0 \pm 0°C temperature, 55-65% relative humidity and 12 h light/12 h dark cycle) for one week for acclimation after their purchase and were fed ICDDR, B formulated rodent food and water *ad libitum*. The Federation of European Laboratory Animal Science Associations (FELASA) guidelines and recommendations were followed to reduce the pain and stress of the experimental rats and were approved by the institutional animal ethical committee.

Acute toxicity study

The median lethal dose (LD₅₀) of the extract in rat was estimated by the up and down method (Bruce, 1985). Doses were adjusted up or down by a constant multiplicative factor (1.5) depending on the previous outcome.

In vivo analgesic activity

The analgesic activity of the methanolic extract of *Z. rugosa* leaves was evaluated using acetic acid-induced writhing method in rat (Sharma et al., 2010). At first, twenty four animals were divided into four groups with six rats in each.

Group I: Treated with vehicle (1% Tween 80 in water, 10 ml/kg body weight p.o.)

Group II: Received Indomethacin (10mg/kg) body weight (p.o.)

Group III: Treated with 200 mg/kg body weight (p.o.) of the extract

Group IV: Treated with 300 mg/kg body weight (p.o.) of the extract.

The test samples and vehicle were administered orally 30 min before intraperitoneal administration of 0.7% v/v acetic acid but Indomethacin (reference drug) was administered orally 15 min before injection of acetic acid. After an interval of 5 min, the mice were observed for specific contraction of body referred to as 'writhing' for the next 10 min. Complete writhing was not always accomplished by the animal; this incomplete writhing was considered as half-writhing. Accordingly, two half-writhing were taken as one full writhing. The number of writhing in each treated groups was compared to that of a control group. Samples having analgesic activity will reduce number of writhes of treated rats. The percent inhibition (% analgesic activity) was calculated by:

$$\% \text{ inhibition} = \frac{A-B}{A} \times 100$$

Where A = average number of writhing of control group; B = average number of writhing of test group.

CNS depressant activity

Open field test

The open field test was carried out to evaluate the effect of extract on locomotor activity of rats by the method described by Gupta et al. (1971). Here, rats were divided into 4 groups of 6 rats each. The control group received 1% Tween 80 in water (10 ml/kg body weight), the standard group received Diazepam (1 mg/kg body weight) and the experimental groups received crude extract at 300 and 500 mg/kg body weight. The floor of an open field was divided into a series of squares each alternatively colored black and white. The apparatus had area of half square meter and a wall of 40 cm height. The number of squares visited by the animals was counted

Table 1. Analgesic effect of methanolic extract of *Z. rugosa* leaves in acetic acid-induced writhing test.

Group	Dose (mg/kg b.w.)	No. of writhing	Percent of inhibition (%)
1% Tween 80 in water (Control)	10 ml/kg	15±.894	
Indomethacin (Standard drug)	10	6.75±.524**	55.20
Leaves extract of <i>Z. rugosa</i>	200	9.92±.585*	33.87
	300	7.22±.785**	51.87

All values are expressed as mean ± SD, (n=6); One way Analysis of Variance (ANOVA) followed by Dunnett's test. *P < 0.05, **P < 0.01, significant compared to control.

Table 2. CNS depressant activity of methanol extract of *Z. rugosa* leaves in open field test in rats.

Treatment	Doses	No. of movement				
		0 min	30 min	60 min	90 min	120 min
1% Tween 80 in water (Control)	10 ml/kg	118.4±2.70	118±2.91	115.4±1.14	117.4±2.61	118±1.58
Diazepam (Standard drug)	1 mg/kg	117.2±2.59	77.6±2.21**	41.8±2.30**	21.8±1.32**	11.6±1.04**
Leaves extract of <i>Z. rugosa</i>	300 mg/kg	118.2±1.92	80.8±2.32**	53.2±0.48**	37.4±1.08**	22.6±2.42**
	500 mg/kg	117.3±2.09	66±1.53**	41±1.10**	19.4±0.14**	12.4±2.00**

All values are expressed as mean ± SD, (n=6); One way Analysis of Variance (ANOVA) followed by Dunnett's test. **P < 0.01, significant compared to control.

for 3 minute, on 0, 30, 60 and 120 min during the study period.

Hole cross test

The method described by Takagi et al. (1971) was implemented for this study. Again 24 rats were equally divided into 4 groups. The control group received 1% Tween 80 in water (10 ml/kg body weight), the standard group received Diazepam (1 mg/kg body weight) and the experimental groups received crude extract at 300 and 500 mg/kg body weight. A steel partition was fixed in the middle of a cage having a size of 30 × 20 × 14 cm. A hole of 3 cm diameter was made at a height of 7.5 cm in the center of the partition. The number of passages of rats through the hole from one chamber to other was counted for a period of 3 minute after 0, 30, 60 and 120 min of oral administration of the extract.

Statistical analysis

All the values in the test are expressed as mean ± standard deviation (SD). The data were statistically analyzed by ANOVA (Analysis of variance) and post-hoc Dunnett's tests with the statistical package for social sciences (SPSS 16.0, USA) program. Dissimilarity between the means of the various groups were measured significantly at p < 0.05 and p < 0.01.

RESULTS

Acute toxicity

Oral administration of graded doses of the methanol

extract of *Z. rugosa* leaves (500 to 5000 mg/kg, body weight) did not cause any death in different dose groups. The LD₅₀ value for oral administration of the plant extract was found to be greater than 5000 mg/kg body weight.

In vivo analgesic activity

In acetic acid induced writhing test, the methanol extract of *Z. rugosa* significantly (p < 0.05 and p < 0.01) reduced the number of writhing movements induced by intraperitoneal administration of acetic acid solution. The dose-dependent inhibition (Table 1) of abdominal constrictions by the methanol extract indicates anti-nociceptive potential of the plant. The exerted inhibition of writhing at a dose 300 mg/kg body weight was close to the standard non-narcotic analgesic drug, Indomethacin.

CNS Depressant Activity

Open field test

The extract was evaluated by open field test to determine the decreasing capability of CNS-locomotor activity in rat model. The extract significantly decreased the locomotor activity in a dose dependant manner (Table 2) and this effect was evident from the initial observation (0 min) period and continued up to 5th observation period (120 min).

Table 3. CNS depressant effect of methanol extract of *Z. rugosa* leaves on hole cross test in rats.

Treatment	Doses	No. of movement				
		0 min	30 min	60 min	90 min	120 min
1% Tween 80 in water (Control)	10 ml/kg	13.67±1.37	14±1.09	13.33±1.03	11.33±1.86	9.17±1.60
Diazepam (Standard drug)	1 mg/kg	10.67±1.03*	5.83±1.33**	5.171±1.52**	4.5±1.52**	3.33±1.03**
Leaves extract of <i>Z. rugosa</i>	300 mg/kg	13.83±1.47	11.5±1.05	9.67±0.816**	8.17±1.47**	7.67±1.03**
	500 mg/kg	11±1.26	9.17±2.56*	7.33±2.42**	5.83±1.72**	3.83±1.17**

All values are expressed as mean ± SD, (n=6); One way Analysis of Variance (ANOVA) followed by Dunnet's test. *P < 0.05, ** P < 0.01, significant compared to control.

Hole cross test

In this test, the extract showed a decrease in locomotion in the test animals. The number of crossing hole from one chamber to another by rat of the control group remained almost steady to slight decrease from 0 minute to 120 minute (Table 3). But the extract at 300 and 500 mg/kg dose showed significant (indicate p-value) gradual decrease of movement from 0 to 120 min. The extract displayed dose dependent activity and maximum depressive effect was observed at fifth (120 min) observation period. Depression produced at 500 mg/kg body weight was found to be close to that of standard drug, Diazepam.

DISCUSSION

The present study was conducted to elucidate analgesic and CNS depressant activities of the methanol extract of *Z. rugosa* leaves. The relatively high oral median lethal dose (LD₅₀) in rat suggests that the extract is relatively non toxic when taken orally (Lorke, 1983; Magaji et al., 2008). Acetic acid-induced abdominal constriction test is used for the evaluation of peripheral analgesic activity (Gene et al., 1998; Faruk et al., 2015). The extract showed significant analgesic activity in acetic acid-induced writhing test in rats. This indicates that the extract possessed peripheral mediated analgesic activity. The abdominal constriction response is thought to involve, in part, the local peritoneal receptors (Bentley et al., 1983; Sani et al., 2013), so the extract may have interfered with these peritoneal receptors to bring analgesia. Acetic acid-induced writhing test has been associated with increase in the levels of prostaglandins E₂ and F_{2α} in peritoneal fluid (Deradt et al., 1980) as well as lipooxygenases (Levini et al., 1984; Sani et al., 2013). So the mechanism of activity of the extract may be linked to cyclooxygenases and/or lipooxygenases.

CNS depressing agents are gaining importance in treating mental disorders like anxiety, dizziness and restlessness. So, we carried out locomotion test to confirm the CNS activity of the methanol extract of the

plant. Increased locomotor activity (by excitatory neurotransmitter) is considered as alertness while decreased locomotor activity (due to inhibitory neurotransmitter) indicates sedative effect (Verma et al., 2010). Our extract had decreased locomotor activity indicating its CNS depressant activity. The CNS depressant activity is observed when the GABAA receptor is activated either by Gamma-amino-butyric acid (GABA) or any other chemical substances. Different anxiolytic, muscle relaxant and sedative-hypnotic drugs exert their action through GABAA receptor. Therefore it is possible that the extract of *Z. rugosa* leaves may act either by potentiating GABAergic inhibition in the CNS via membrane hyperpolarization or may be due to direct activation of GABA receptor (Kolawole et al., 2007). Many research results showed that plant containing flavonoids, saponins and tannins are useful in many CNS disorders and many flavonoids and neuroactive steroids were ligand capable for the GABAA receptors in the central nervous system. This led to the assumption that they can act as benzodiazepine like molecules (Bhattacharya and Satyan, 1997).

Conclusion

Based on the results of the present study, we conclude that the methanolic extract of *Z. rugosa* leaves possesses remarkable dose dependent analgesic and CNS depressant activity. However, further studies are indispensable to examine the underlying mechanisms of such CNS effects and to isolate the active compounds responsible for these pharmacological activities.

Conflict of interest

The authors have not declared any conflict of interest

REFERENCES

- Abhimany Y, Pratiksha S (2010). Analgesic and anti-inflammatory activities of *Zizyphus rugosa* root barks. J. Chem. Pharmaceu. Res. 2: 255-259.

- Bhattacharya SK, Satyan KS (1997). Experimental methods for evaluation of psychotropic agents in rodents: I--Anti-anxiety agents. *Indian. J. Exp. Biol.* 35: 565-575.
- Bruce RD (1985). An up- and down procedure for acute toxicity testing. *Fundam. Appl. Toxicol.* 5: 151-157.
- Devraj A, Karpagam (2011). Evaluation of anti-inflammatory activity and analgesic effect of *Aloe vera* leaf extract in rats. *Int. Res. J. Pharma.* 2(3):103-110.
- Faruk MA, Khan MF, Mian MY, Rahman MS, Rashid MA (2015). Analgesic and anti-diarrheal activities of *Aganosma dichotoma* (Roth) K. Schum. in swiss-albino mice model. *Bang. Pharm. J.* 18(1): 15-19.
- Gawande RK, Tare HL, Shende VS, Bongirwar AA, Deore SR, Dama GY (2011). Anxiolytic and CNS depressant activity of extracts obtained from seeds of *Zizyphus rugosa*. *Int. J. Cur. Pharmaceu. Clin. Res.* 1:21-32.
- Gene RM, Segura L, Adzet T, Marin E, Inglesias J (1998). Heterotheca inuloides: anti-inflammatory and analgesic effects. *J. Ethnopharmacol.* 60: 157-162.
- Bakshi GDN, Sensarma P, Pal DC (1999). A lexicon of medicinal plants in india. 1st Edition, Kolkata, India Naya Prokash. 1: 205-206.
- Bentley GA, Newton SH, Starr J (1983). Studies on the anti-nociceptive action of agonist drugs and their interaction with opioid mechanisms. *Br. J. Pharmacol.* 32: 295-310.
- Deradt R, Jougney S, Delevalcee F, Falhout M (1980). Release of prostaglandin E and F in an algogenic reaction and its inhibition. *Eur. J. Pharmacol.* 51:17-24.
- Ferdous MR, Rashid RB, Sikder MA, Aktar F, Rashid MA (2012). Preliminary in vitro biological and phytochemical screenings of *Parmentiera cereifera* Seem. *Bang Pharm. J.* 15(2): 103-106.
- Gupta BD, Dandiya PC, Gupta ML (1971). A psychopharmacological analysis of behavior in rat. *Jpn. J. Pharmacol.* 21:293-298.
- Khan MF, Khan ZI, Uddin MR, Rahman MS, Rashid MA (2015). n vivo hypoglycemic and alloxan induced antidiabetic activity of *Xeromphis uliginosa* Retz. *Afr. J. Pharm. Pharmacol.* 9(11): 363-366.
- Kekuda TR, Vinayaka KS, Mallikarjun N, Bharath AC, Shailendra B, Rakesh MC, Vinod HR (2011). Antibacterial, insecticidal and free radical scavenging activity of methanol extract of *Zizyphus rugosa* Lam. fruit pericarp. *Pharmacog. J.* 2: 65-69.
- Kolawole OT, Makinde JM, Olajid OA (2007). Central nervous depressant activity of *Russelia equisetiformis*. *Nig. J. Physiol. Sci.* 22: 59-63.
- Lorke D (1983). A new approach to acute toxicity testing. *Arch. Toxicol.* 54:275-287.
- Magaji MG, Anuka JA, Abdu-Aguye I, Yaro AH, Hussaini IM (2008). Preliminary studies on anti-inflammatory and analgesic activities of *Securinega virosa* (Euphorbiaceae) in experimental animal models. *J. Med. Plant. Res.* 2(2):039-044.
- Pandey VB, Tripathi YC, Devi S, Singh JP, Shah AH (1988). A cyclopeptide alkaloid from the bark of *Zizyphus rugosa*. *Phytochem.* 27(6):1915-1918.
- Prashith Kekuda TR, Raghavendra HL, Vinayaka KS (2011). Evaluation of pericarp and seed extract of *Zizyphus rugosa* Lam. for cytotoxic activity. *IJPBA.* 2:887-890.
- Rahman MS, Begum B, Chowdhury R, Rahman KM, Rashid MA (2008). Preliminary cytotoxicity screening of some medicinal plants of Bangladesh. *Dhaka Univ. J. Pharm. Sci.* 7(1):47-52.
- Rambabu P, Venkata Ramana K, Ganapaty S (2011). Anti bacterial and anti fungal potential of roots of *Zizyphus rugosa*. *IJPRD,* 3: 85-93.
- Sani YM, Musa AM, Yaro AH, Amoley A, Magaji MG (2013). Phytochemical screening and evaluation of analgesic and anti-inflammatory activities of the methanol leaf extract of *Cissus polyantha*. *J. Med. Sci.* 13(8): 824-828.
- Sarker M, Das SC, Saha SK, Mahmud ZA, Bachar SC (2012). Analgesic and anti-inflammatory activities of flower extracts of *Punica granatum* Linn. (Punicaceae). *J. Appl. Pharma. Sci.* 2(4): 133-136.
- Sharma A, Bhatial S, Kharyaz MD, Gajbhiye V, Ganesh N, Namdeo AG (2010). Anti-inflammatory and analgesic activity of different fractions of *Boswellia seratta*. *Int. J. Phytomed.* 2:94-99.
- Singh A, Pandey MB, Singh S, Singh AK, Singh JP (2009). Flavonoids of *Zizyphus rugosa*. *J. Indian. Chem. Soc.* 86(2):177-178.
- Takagi K, Watanabe M, Saito H (1971). Studies on the spontaneous movement of animals by the hole cross test: Effect of 2-dimethylaminoethane, its acylates on the central nervous system. *Jpn. J. Pharmacol.* 21:797-810.
- Verma A, Jana GK, Sen S, Chakraborty R, Sachan S, Mishra A (2010). Pharmacological evaluation of *Saraca indica* leaves for central nervous system depressant activity in mice. *J. Pharm. Sci. Res.* 2(6):338-343.

Full Length Research Paper

Development of nano particle encapsulated pemulen gel for aceclofenac topical delivery

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Received 24 May, 2016; Accepted 7 September, 2016

Management of rheumatic disorders are limited by the inability to provide long-term drug delivery due to the irritation to gastric mucosa and poor availability of drugs, which may be overcome by prolonging the drug release. The nanostructured carriers (NP)based gel was developed as potential topical system for aceclofenac topical delivery. In the present study, chitosan-sodium alginate nanoparticles were investigated using a novel gel base for the prolonged topical delivery NSAID aceclofenac. A modified ionotropic gelation method was used to produce aceclofenac loaded nano particulate systems. Drug content, particle properties such as size, size distribution and zeta potential were determined. The nano particulate system was encapsulated using a novel hydrophilic polymer pemulen in different proportions. The nanoparticles remained within the colloidal range and it was uniformly dispersed in pemulen gel preparation. The gels were evaluated for spreadability, extrudability and in vitro, in vivo studies. The optimized formulations were characterized by FT-IR, DSC. In conclusion, nanoparticle based gel could be a promising vehicle for topical delivery of aceclofenac and the novel gel base pemulen can be used for the incorporation topical drugs.

Key words: Pemulen, aceclofenac, sodium alginate, transdermal, chitosan, nanoparticle, topical delivery, gel.

INTRODUCTION

Aceclofenac is a cyclooxygenase inhibitor used in the treatment of osteoarthritis, rheumatoid arthritis and in management of acute pain in adults. It shows high anti-inflammatory and analgesic activities with the incidence of gastric side effects and high therapeutic index (Fei et al., 2012). The mean plasma elimination half-life is around 2 h and is administered at a dose of 100 mg twice daily. The shorter biological half-life and dosing frequency more than once a day for prolonged period results in

gastric irritation (Batheja et al., 2013).

Nanoparticles are novel drug delivery systems having the capability to release the drug at an optimum rate at the desired site of action. Nano particular formulations provide the liberty to use a wide range of polymers like synthetic, natural, biodegradable and non-biodegradable polymers. (Harshad et al., 2013). Chitosan and alginate are two biopolymers that have received much attention and have been extensively studied for such use

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Chitosan being a cationic polymer has been used for the production nanoparticles by ionotropic gelation with negatively charged polymers and there are many chitosan-polyanion complexes that have been investigated as drug delivery systems for drugs, proteins with encouraging results (Shahin et al., 2011)

Nano particles are promising delivery carriers that have been utilized for formulation and delivery of various drugs. For topical administration, they are usually incorporated into gel or cream to increase their residence time, their stability and characteristics. The topical use of nanoparticles improves the drug bioavailability by prolonging the residence time of drugs applied topically or by enhancing the passing of drugs through the epithelial cells by opening the tight junctions between epithelial cells and also to reduce the side effect of the drug (Luigi et al., 1995; Alessandro et al., 2015).

Pemulen is a polymeric emulsifier composed of a block copolymer consisting of a poly acrylic acid similar to the Carbopol used to make aqueous and solvent gels. Pemulen is part of a class of copolymers, referred to as acrylate/C10-30 alkyl acrylate cross polymers which is cross linked with a long-chained methacrylate having a lipophilic regions as the methacrylate as well as hydrophilic regions composed of the acrylic acid. Cross-linked polymers can swell in water up to 1,000 times their original volume to form a gel (soomneka et al., 1995).

Pemulen polymers deposit an occlusive layer on the skin, delivering the topical medication in the form of low-irritancy formulation with elegant skin feel. Pemulen polymer also be used for high-clarity topical gels (Isabelle, 2014; Elgadir et al., 2015). The current study aimed at developing nano particulate topical formulation of aceclofenac using a novel polymer pemulen as a gel base for topical delivery.

MATERIALS AND METHODS

Aceclofenac was obtained as a gift sample from MSN Lab Pvt Ltd. Hyderabad and pemulen was gift from Strides Lab, Bangalore. Methylparaben, Propylparaben and polyethylene glycol was procured from Merck Chemicals, Mumbai. Sodium Alginate was Procured from SD Fine Chemicals Mumbai and chitosan was collected as a gift sample from Central Institute of Fisheries Technology, Kochin. All the solvents used were analytical grade and double distilled water was used for experimental work.

Experimental

The pemulen based transdermal delivery system was developed by encapsulating sodium alginate based aceclofenac nanoparticles in pemulen gel base. Sodium alginate nanoparticles were prepared employing ionotropic pregelation method and further they were encapsulated in a gel using pemulen as a base and the efficiency of gel base in delivery of aceclofenac was evaluated.

Preparation and evaluation of sodium alginate nanoparticles

Sodium alginate nanoparticle formulated employing two-step

process based on ionotropic pre-gelation method with modifications from ideal preparation. The nanoparticles of aceclofenac were prepared based pre-gelation of poly anion with calcium chloride followed by poly cationic crosslinking technique.

The nanoparticles were developed using 0.0063% w/v sodium alginate solution. Drug was incorporated at the rate 1 mg/mL to sodium alginate solution in step one itself, before adding to calcium chloride for gelation. About Calcium chloride solution (7.5 mL) was added drop wise to sodium alginate solution (117.5mL, 0.0063% w/v) to induce gelation. It was stirred continuously for 60 min to get the alginate pre gel. Then different concentrations of 25 mL of chitosan solution was added drop wise to the pre gel along with constant stirring for 90 min. Nanoparticles were concentrated by centrifugation at 11,500 rpm for 40 min. Nanoparticles thus formed were analyzed for particle size (Soomneka et al., 1995).

The volumes of the acetic acid and sodium hydroxide were selected in a way to adjust the final pH at around 5.9 to 6, to ensure the solubility of both aceclofenac and chitosan and the stability of the prepared nanoparticles (Monika Schäfer-Korting et al., 2007; Lippacher et al., 2001).

Formulation of aceclofenac pemulen gel

Pemulen at different concentration (0.5, 1 and 2% w/w) were used to formulate different topical formulation of aceclofenac Different formulations of pemulen based aceclofenac gel is given in Table 1. Pemulen was dispersed in water with preservatives methyl paraben and propyl paraben which were dissolved in freshly boiled and cooled water.

Aceclofenac was dissolved in propylene glycol and glycerin mixture and it was added to the preservative mixture and stirred well. Then the aqueous solution was added to the pemulen dispersion with stirring to get the uniform gel consistency. The pH of gel was adjusted within the range of 6.8 to 7.2 by adding triethanolamine. The uniform dispersion was packed in collapsible tubes for further studies (Simovic et al., 2007).

Preparation of nano particle encapsulated pemulen gel

The optimized batch of the nano particle dispersion dispersed into different concentration of the pemulen gel. Pemulen was soaked in preservative solution for overnight as specified previously. The nanoparticles equivalent to 100 mg of aceclofenac was dispersed in glycerin propylene glycol mixture and mixed with pemulen gel to get the uniform consistency of pemulen gel. Later the pH was adjusted with tri ethanolamine. Three batches of pemulen gels were prepared and subject to physical and chemical analysis.

Evaluation studies

Zeta size

The average particle size and poly dispersity index of the nanoparticles were determined by photon correlation spectroscopy using a Zeta sizer DTS version 5.03 (Malvern Instrument, Worcestershire, England). The samples of nanoparticles dispersions were diluted to four times their volume with 0.05 M NaCl. The particle size is represented by the average (diameter) of the Gaussian distribution function in the logarithmic axis mode.

Zeta potential

Zeta potential is an abbreviation for electro kinetic potential in colloidal systems. Zeta potential is electric potential in the interfacial

Table 1. Formulations of nanoparticle encapsulated aceclofenac gel.

Particles	AG1	AG2	AG3	AG4	NPG1	NPG2	NPG3	NPG4
Aceclofenac	1%	1%	1%	1%	1%	1%	1%	1%
Pemulen (%W/W)	0.5	1	1.5	2	0.5	1	1.5	2
Tri ethanolamine	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05
Propylene glycol	1.5	1.5	1.5	1.5	1.5	1.5	1.5	1.5
glycerin(ml)	1.5	1.5	1.5	1.5	1.5	1.5	1.5	1.5
Methyl paraben	0.04	0.04	0.04	0.04	0.04	0.04	0.04	0.04
Propyl paraben	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01
Purified water q.s up to (g)	10	10	10	10	10	10	10	10

AG, Aceclofenac gel; NPG, nanoparticle gel.

double layer at the location of the slipping plane versus a point in the bulk fluid away from the interface. The surface charge (Zeta potential) was determined by measuring the electrophoretic mobility of the nanoparticles using a Malvern zeta sizer (Malvern instrument, UK). Samples were prepared by diluting with distilled water.

Physical analysis of pemulen gel

The following physical analysis was carried out for pemulen based aceclofenac and its nanoparticle encapsulated topical formulations. The organoleptic features of the pemulen gels were examined at the same temperature, lighting and packaging condition to assess variation in appearance, phase separation and color.

pH measurements

One gram of each pemulen gel formulation was weighed and dispersed with 25 ml of distilled water. After homogenization, the pH of the sample was measured with meter. The test was conducted as triplicate.

Spreadability of gel

For gel dosage form good spreadability value is one of the important properties. Spreadability gives indication of gel ability to spread on skin part. Spreading value decide the therapeutic efficiency of gel. Spreadability apparatus contains wooden block having two glass plates. Initially gel sample was placed between the two glass plates. Weight near about 300 g was putted on top plate which expelled the air and form uniform gel layer. Afterwards 100 g weight was put to drag top plate by 10 cm using string attached to hook. Time required to move upper plate by 10 cm distance was noted, lesser the time required for dragging the upper plate better is the spreadability value. Spredability value was determined with the help of formula:

$$S = M \times L / T$$

Where, S is the spreadability value, L is the length of the glass slide, M is the weight tied to the upper plate, and T is the time taken to separate the glass slides. The test was conducted as triplicate for all formulations.

Extrudability

A good gel formulation should extrude easily from the container. In

this test, a closed collapsible tube containing 10 g gel was passed firmly at crimped end. When the cap was removed, gel extrudes until pressure was dissipates. The weight in grams required to extrude 0.5 cm ribbon of gel in 10 s was determined. The results for each formulation were recorded as extrusion pressure in grams.

In-vitro drug diffusion study

The diffusion study of the nanoparticle gel and conventional gel using pemulen were carried out in Franz diffusion cell. The cellophane membrane was soaked in phosphate buffer for 24 h and it was mounted on the diffusion cell for further studies. Gel sample containing 100 mg of aceclofenac was taken on the semi-permeable dialysis membrane presoaked overnight in the freshly prepared diffusion medium. The donor compartment was kept in contact with a receptor compartment and the temperature was maintained at $37 \pm 0.1^\circ\text{C}$. The buffer solution was kept on the receptor side. At time intervals of 0.5, 1, 2, 3, 4, 5 and 6 h, 2 ml of sample was withdrawn and replaced by fresh medium and the drug concentration on the receptor fluid was determined spectro photometrically at 248 nm against appropriate blank (Sangvi et al., 2007).

Ex-vivo diffusion study

Ex vivo diffusion study for nanoparticle gel and conventional gel were performed on the goat skin using Franz Diffusion cells. Fresh goat hairless skin was obtained from a local slaughter - house. The skin was separated by removing the underlying fat and loose tissues. The membrane was washed with distilled water and then with 0.1 N NaoH and then wrapped in aluminium foil. The film was stored in freeze until further use. For ex vivo permeation studies, skins were allowed to hydrate for 1 h with phosphate buffer before being mounted on the Franz diffusion cell with the stratum corneum (Sharma et al., 2011).

Drug release study was performed on the excised skin using Franz diffusion cell. The skin was cleaned with water and then swabbed with cotton immersed in phosphate buffer. The gel formulations loaded with 10 mg of aceclofenac and its equivalent nanoparticles gel were placed on the excised skin. Stratum corneum was placed towards donor compartment and dermal towards receptor compartment. Phosphate buffer solution (pH 6.8) used as diffusion medium ($37 \pm 0.5^\circ\text{C}$). At time intervals of 0.5, 1, 2, 3, 4, 5, 6 and 12 h and replaced by fresh medium 2 ml sample were withdrawn and drug content was determined by UV spectrophotometer. For determination of drug deposition diffusion cell was dismantled, skin was removed. Excess drug present on the skin was removed. Afterwards skin was cut into small pieces which

were placed in methanol undergo sonication for 1 h. Drug deposition in the skin was checked by using UV spectrophotometer.

Characterization studies

Fourier transform infra-red spectroscopy

About 5 mg of sample was mixed thoroughly with 100 mg of KBr IR powder and compacted under vacuum at a pressure of about 6000 kg/cm² for 3 min. The resultant disc was mounted in a suitable holder in a Shimadzu model 8033 IR spectrophotometer and the IR spectrum was recorded from 4000 to 625 cm⁻¹ in a scan time of 12 min. The resultant spectra were compared for any spectral changes. The resultant spectra were compared for any spectral changes.

Differential scanning calorimetry

Differential scanning calorimetric thermo grams of sample were recorded in Mettler Toledo cu and k irradiation, analyzer equipped with a monitor and printer. The instrument was calibrated with indium as a standard. Accurately weighed 2.5 mg of samples were placed in open flat bottom, pierced aluminium sample pan. The melting point, peak maxima, appearance of any new peak and peak shape was noted.

Anti-inflammatory activity

The anti-inflammatory activity of the nanoparticle encapsulated pemulen gel was assessed employing carrageenan induced paw oedema method. Adult male Wistar rats, weighing between 128 to 160 g, maintained in standard laboratory conditions, at temperature 25 ± 1°C and relative humidity 55 ± 5%. These rats are randomly divided into 3 groups with 6 in each group.

Experimental Procedure: Wister rats were weighed and marks were made on right hind paw behind tibiotarsal junction. Each rat was placed in an observation chamber for 20 min to minimize stress related behaviors. Inflammation was induced by injecting 0.1 ml of 1% w/v carrageenan solution dispersed normal saline subcutaneously into the sub-plantar surface of the right paw of the rat. After one hour about 0.5 g of the gel formulations of plain aceclofenac and nanoparticle encapsulated gel was gently rubbed onto the plantar surface of the right hind paw 50 times with the index finger. All rats were subsequently returned to the observation chamber. The formulation was given as per the following groups.

Group - I: Control (Inflamed, treated with gel base only).

Group - II: Standard (Inflamed, treated with the plain aceclofenac gel)

Group - III: Sample (inflamed, treated with the nanoparticle encapsulated pemulen gel).

The inflammatory response was assessed by measuring the volume of the paw at different periodical intervals after carrageenan administration, using plethysmometer.

The percentage inhibition of paw volume for each rat in treated groups was calculated and expressed as mean ± SD percent inhibition of paw volume. The single ANOVA test was applied to test the significance.

RESULTS AND DISCUSSION

The preparation of sodium alginate aceclofenac

nanoparticles was based on ionotropic gelation process using aqueous phases of the poly cationic chitosan and poly anionic alginate. The optimized nanoparticles of aceclofenac were found to be fine, spherical and free flowing powders. The optimized nanoparticles showed a drug content in the range of 90.76±0.55 to 99.45 ±0.55%. The particle size varied from 287.89 to 356.87 nm. Highest encapsulation efficiency of 87.32% and a smaller particle size particle size of 187.2 nm were taken as criteria for selection of optimized formulation.

Zeta size

Zeta size distribution of the optimized chitosan-alginate nanoparticles given in Figure 1. The particle size distribution of the prepared nanoparticles was determined by using a zeta sizer. The results revealed that the particle size distribution was found to be uniform and the size was found to increase with addition of chitosan.

The particle diameter (z-average) for the optimized chitosan-alginate nanoparticles was approximately 187.4 nm (Figure 1). It is noteworthy that the hydrodynamic diameter of the particles measured by light scattering is smaller than the size estimated from microscopy particularly because of high swelling capacity of chitosan-alginate nanoparticles.

Zeta potential

The Zeta potential of optimized aceclofenac loaded alginate chitosan nanoparticles is given in Figure 2. For the optimized nano formulation, value of zeta potential was found to be -35.6 mV. This relatively high positive value was due to cationic nature of Chitosan and Alg/Chi ratio. The net positive zeta potential indicates the presence of free surface amino groups on the nanoparticulate delivery system. It indicates that the synthesized drug loaded nanoparticles could be expected to be stable for long time.

The stability of colloidal systems is directly related to the magnitude of their zeta potential. In general, if the value of the particle zeta potential is large, the colloidal system will be stable. Conversely, if the particle zeta potential is relatively small, the colloidal system will agglomerate. The surface charge of the particles is of substantial importance in all the production steps of these particles because the efficiency of the delivery system is directly related to the establishment of electrostatic interactions.

Evaluation of pemulen gels

All topical pemulen based formulation aceclofenac gels were clear, homogenous without any phase separation. The gels were transparent, non-greasy without any gritty

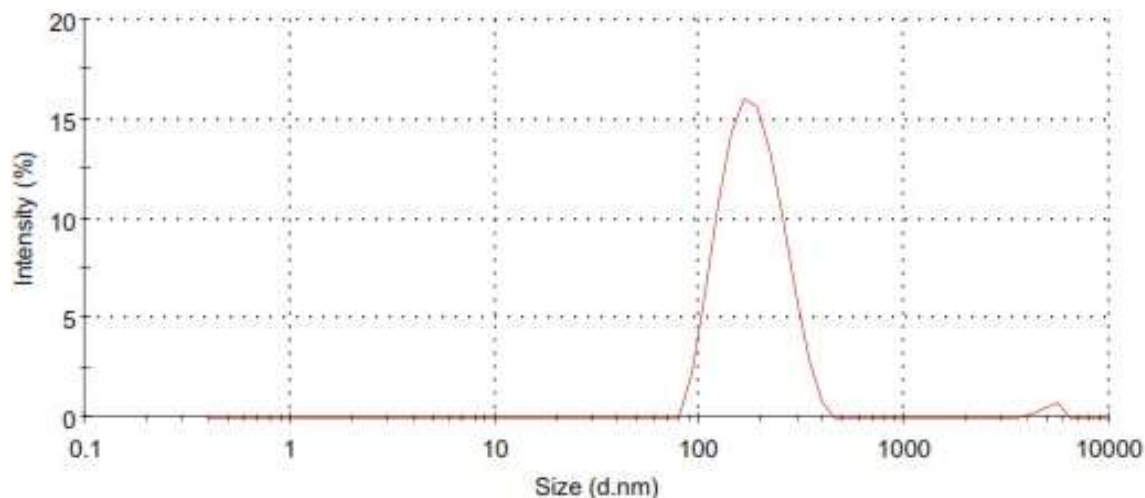


Figure 1. Zeta size distribution of optimized aceclofenac loaded chitosan-alginate nanoparticles.

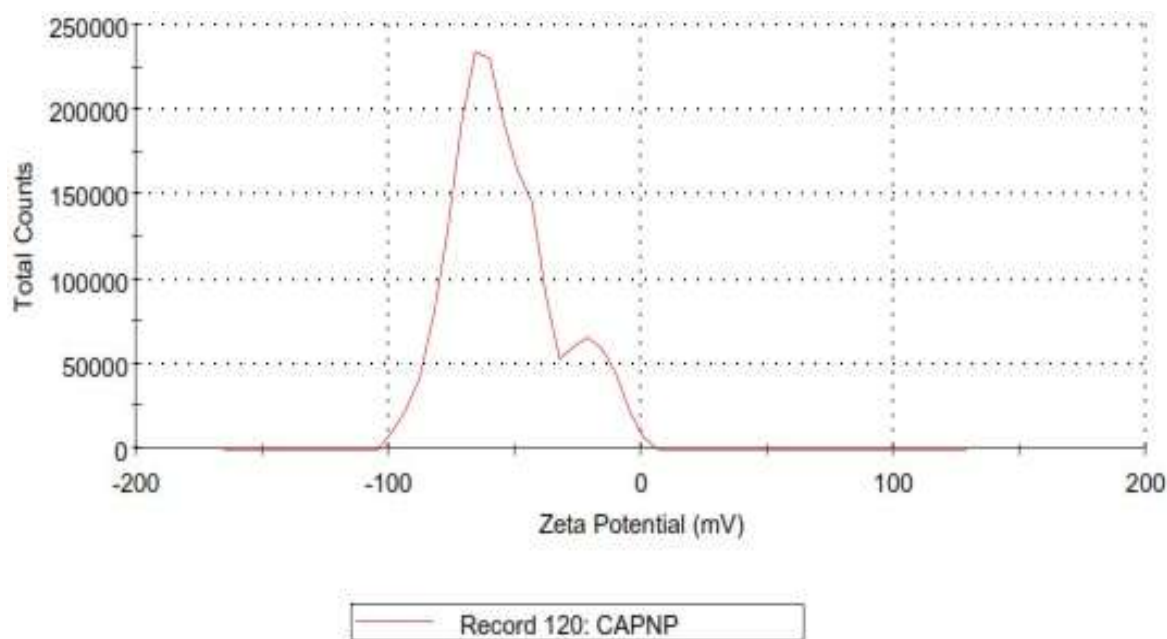


Figure 2. Zeta potential of optimized aceclofenac chitosan-alginate nanoparticles.

particles. Spreadability data indicate that the gel is easily spreadable by a small amount of shear. Consistency reflects the capacity of the gel to get ejected in uniform and desired quantity when the tube is squeezed. The gel formulations were homogeneous in texture and fell within a pH range of 6.8 to 7.4 which is within the normal skin pH.

The aceclofenac and its nanoparticle encapsulated gels were smooth, elegant and transparent. The gel showed homogenous gel without any grittiness. The nanoparticle encapsulated gel showed uniform distribution of nanoparticles.

pH Determination

The pH values of all developed formulae was in range 6 to 7 which is considered acceptable to avoid the risk of irritation upon application to the skin. The results propose that the nano particle-based semi-solid formulation is acceptable for topical application.

Drug content

Drug Content determination results are Tabulated in Table 2 after various formulation of aceclofenac gel the

Table 2. Physico chemical evaluation of different formulations of pemulen gel (n=6) (AG-ACECLOFENAC and NPG-NANOPARTICLE GEL).

Formulation	Drug content	Spreadability	pH	Homogeneity consistency
AG ₁ (0.5%)	99.8±0.32	8.5±0.31	7.3±0.3	Good
AG ₂ (1%)	97.5±0.45	8.6±0.43	7.1±0.2	Very good
AG ₃ (1.5%)	97.5±0.45	8.4±0.39	7.2±0.2	Very good
AG ₄ (2%)	94.89±0.89	7.6±0.34	6.8±0.4	Good
NPG ₁ (0.5%)	96.90±0.73	6.8±0.46	7.4±0.3	Poor
NPG ₂ (1%)	97.90±0.56	5.4±0.31	6.8±0.2	VERY GOOD
NPG ₃ (1.5%)	96.89±0.82	4.7±0.33	6.4±0.1	Good
NPG ₄ (2%)	94.89±0.20	4.4±0.93	6.0±0.1	Poor

Table 3. Drug release kinetics of optimized-pemulen aceclofenac gels.

Gels	Zero order	Peppas	Higuichi	First Order
Aceclofenac gel (AG3)	0.83746	0.9797	0.9782	0.8239
Nanoparticle pemulen gel (NPG2)	0.9867	0.9975	0.9566	0.9847

drug content of the formulated gel was estimated and the results were in the official limits with range of 9.5 to 9.99 mg/g gel. The drug content determination also showed that the drug was uniformly distributed throughout the gel.

Extrudability and spreadability

The extrusion of gel is important parameter during application and for the patient compliance. Extrudibility of gel formulations with high concentration of gelling agent was found satisfactory while with low concentration of gelling agents good extrudability was observed. Spreadability plays an important role in patient compliance and help in uniform application of gel to the skin. Pharmaceutically acceptable gel takes less time to spread and will have high spreadability. The spreadability of formulation was decreased as the concentration of Pemulen was increased. The spreadability of formulation was decreased as the concentration of Pemulen was increased it was assumed that the increased viscosity at higher concentration of pemulen was responsible for decreased spreadability.

In vitro drug release studies

The *in vitro* drug release studies of the aceclofenac gel containing different proportions of pemulen is shown in Figure 3. As the concentration of pemulen in the dispersion increased, the dissolution rate also decreased. Among all the formulation containing 1% of pemulen showed complete release of drug in 30 min whereas increased proportions of pemulen showed slower and incomplete drug release. This could be attributed to the

fact that very high quantity in the dispersion may be responsible for impeding the drug diffusion through the dispersion matrix thus bringing about a reduction in drug release profile from the formulated system.

The comparative profile of optimized pemulen with its nanoparticulate dispersion shows that the nanoparticles showed delayed release for a period of 6 hs shown in Figure 4. It was presumed to be due to controlled, slower release of the drug from nano particulate particle. The plain aceclofenac nano particle showed more than 75% drug diffused while the nano particulate based gel showed decreased drug release. While the gel formulation containing 2% of pemulen showed decreased drug release than other formulation which was assumed to be due to the high viscosity of gel.

Kinetics of drug release

The comparative release constants of aceclofenac and nano particle encapsulated gel are given in Table 3. The release kinetics data indicated that the release of drug from nano particle best fits to zero order release model because the correlation coefficient values were higher in case of zero order equation and the release from gel NPG2 fits to Higuchi model. The release rate is independent of the concentration of the drug. The release exponent value of Korsmeyer-Peppas signifies that the mechanism of drug release from both gel and nanoparticle gel follows anomalous diffusion.

Ex vivo drug permeation studies

Ex vivo permeation of aceclofenac from nano particle

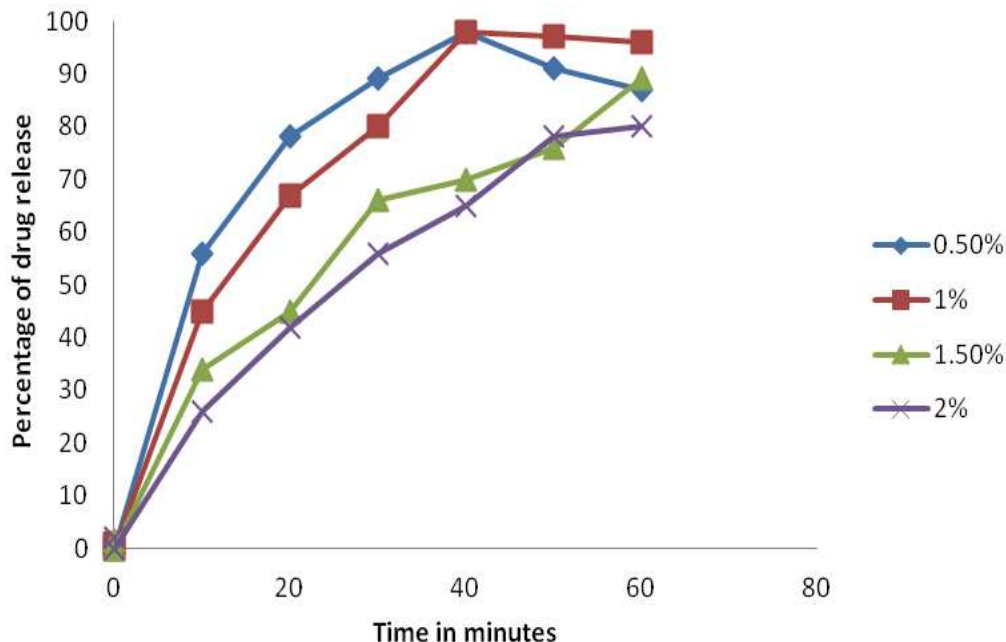


Figure 3. *In vitro* drug release studies of different formulations of aceclofenac pemulen gel.

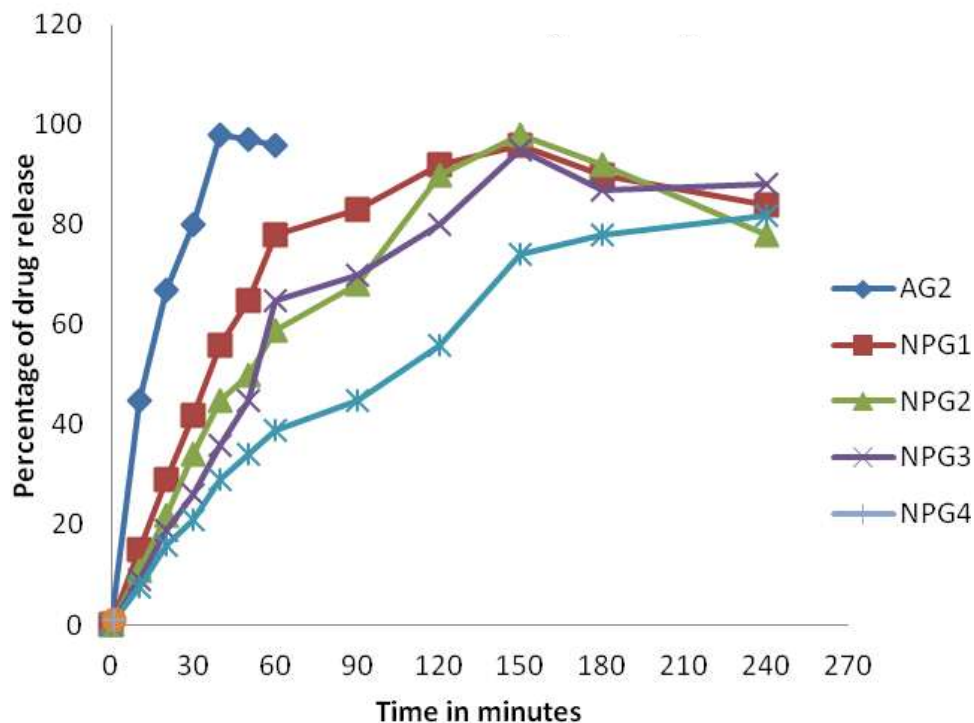


Figure 4. Comparative drug release studies of the pemulen based nanoparticle gel.

encapsulated gel through excised rat skin was assessed and correlated with plain aceclofenac gel.

The amount of drug permeated through rat skin from aceclofenac nanoparticle incorporated gels is given in

Figure 5. Nano particle dispersion gel exhibited the greatest cumulative amount of drug permeation in 12 h. The cumulative amount of drug permeated at the end of 12 h was found to be greater for aceclofenac nanoparticle

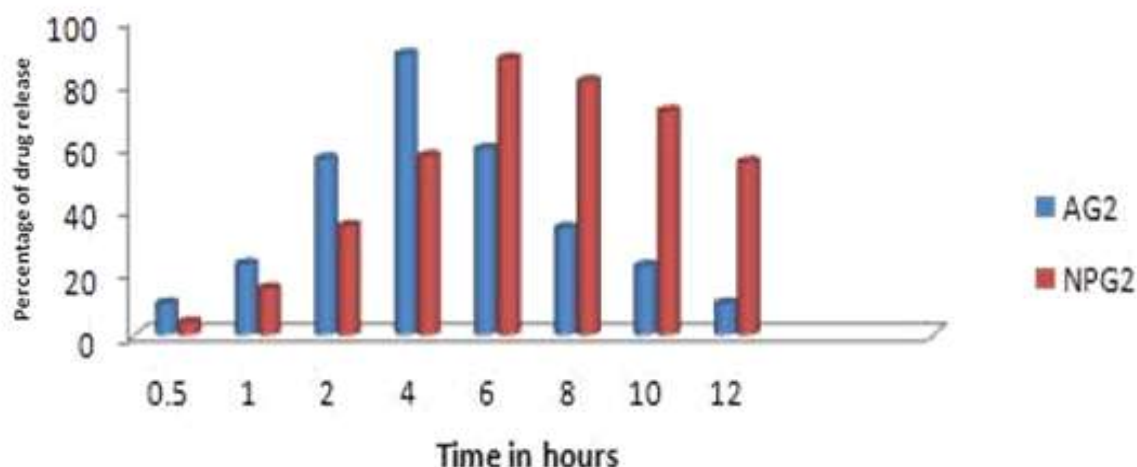


Figure 5. *Ex vivo* release studies of nanoparticle incorporated aceclofenac pemulen gel.

enriched pemulen gels than the aceclofenac gel nanoparticle encapsulated pemulen gels released drug slowly when compared with plain ACN gels accounted for by the time drug takes to diffuse through the gel. The slower release of drug from nanoparticle gels maintained the drug concentration for longer period. The formulations possessed a sustained drug release, but the sustained effect was more pronounced with nanoparticle enriched gel formulations.

The amount of ACN penetrated into dermis from NP gel at 12 h was nearly 2 fold that of plain aceclofenac gel. The small size and close interaction between NPs and the stratum corneum are the possible reasons for increase in the drug amount penetrating into the viable skin. The increase amount of drug in the dermis is also related to the occlusion properties. It was assumed that hydration of stratum corneum can increase, which can facilitate drug penetration of transdermal drug delivery.

Characterization studies

FTIR spectra of aceclofenac encapsulated pemulen gel

The FTIR spectra of aceclofenac (Figure 6) showed distinct sharp peaks at 3317 and 3272 indicate the presence of a primary amine and broad peaks near 2919 including 1921 may be due to CH stretching of CH₂ groups, carbonyl group vibration at 1770 and 1716. Peaks at 1589, 1577 and 1508 indicates the presence of C=C ring stretching. The characteristic peak of 1723 was observed due to the pemulen. All these principal IR peaks of aceclofenac and pemulen were also observed in FTIR spectrum of aceclofenac gel. No extra peaks were found in spectrum of aceclofenac gels indicating absence of interaction or instability in gel formation.

DSC studies nanoparticle encapsulated gel

The DSC of aceclofenac showed a sharp endothermic peak at 149.40°C corresponding to the melting point of aceclofenac. The DSC of aceclofenac and its optimized nanoparticle gel is given in Figure 7. The DSC scan of physical mixture also showed a sharp melting at 152.87 due to the melting of pemulen and a melting point at 272 is due to melting of sodium alginate and at 122.7°C due to chitosan melting. The DSC of optimized formulation with no additional peak indicating that the aceclofenac did not undergo any crystal modification or degradation. The gel formulation showed a melting point at 137°C indicates slight alteration of melting point it was assumed that the interaction of sodium alginate with chitosan was responsible for alteration of melting point. The intensity was decreased due to the dilution in pemulen gel.

As the DSC scan does not showed significant changes it can be concluded that the aceclofenac does not showed any crystalline changes.

Anti-inflammatory activity aceclofenac nanoparticle gel

Anti-inflammatory activity of aceclofenac and its sodium alginate nanoparticles gel in pemulen is reported in Figure 8. The anti-inflammatory activity of nanoparticles in comparison with pure drug was evaluated on the basis of ability to inhibit the edema produced in hind paw after challenging with carrageenan.

The sodium alginate nanoparticles of aceclofenac gel showed delayed action as compared to pure drug. A peak of inhibition of 31.6 and 54.3% was observed after 6 h for plain drug and nanoparticles respectively. The difference was significant showing significance with a *p* value less than 0.005% indicating that the nanoparticles

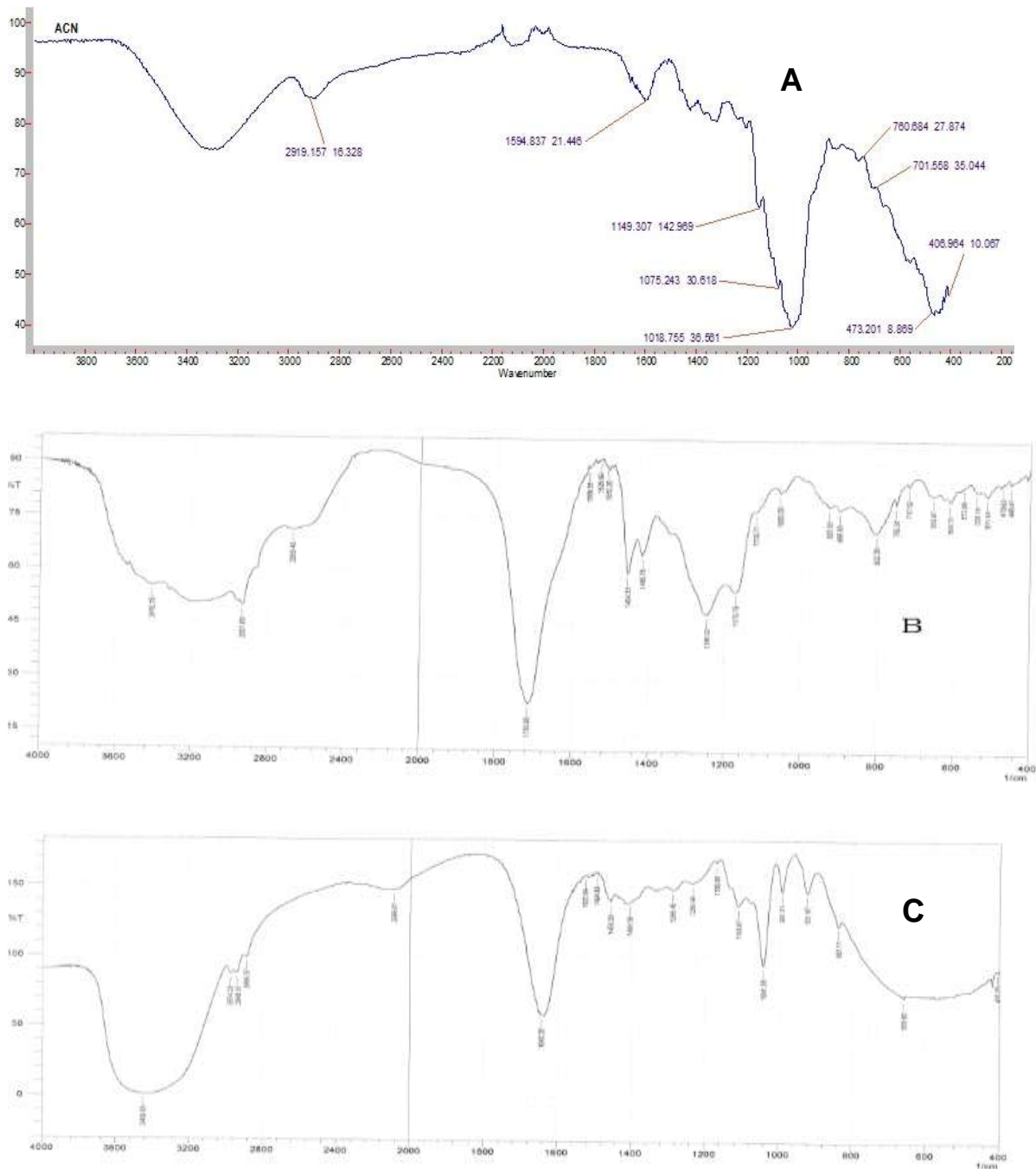


Figure 6. FTIR spectra of (a) aceclofenac, (b) gel physical mixture, and (c) nanoparticle gel.

gel of aceclofenac showed an improvement in anti-inflammatory activity for prolonged period on topical application.

Conclusion

The research on the formulation of pemulen based nanoparticle encapsulated aceclofenac gel indicates that the nanoparticulate encapsulated gel can be an effective

formulation strategy for the management of rheumatoid patients. The novel gelling agent pemulen can be used as an effective formulation ingredient in topical preparations.

Conflict of Interests

The authors have not declared any conflict of interests.



Figure 7. DSC of aceclofenac and its sodium alginate nanoparticles encapsulated gel.

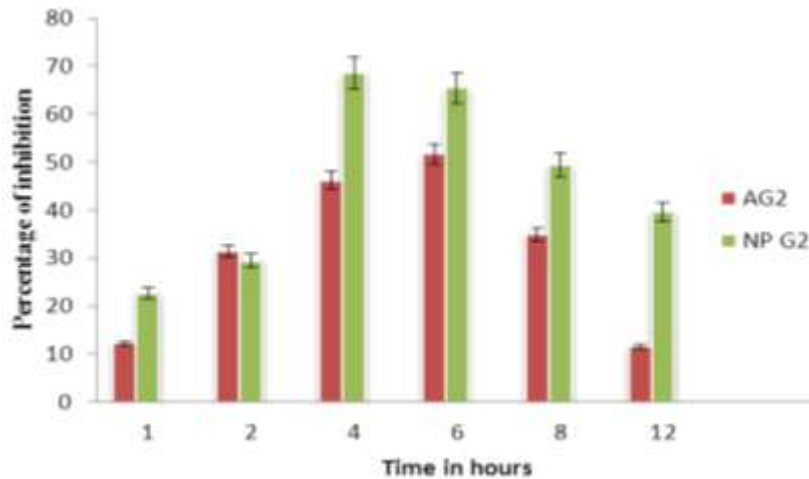


Figure 8. Comparative anti-inflammatory activity of aceclofenac nanoparticle encapsulated gel with plain aceclofenac gel.

REFERENCES

- Alessandro FM, Monteiro JP, Bonafé EG, Gerola AP, Silva CTP, Girotto EM, Rubira AF, Muniz EC (2015). Bactericidal activity of hydrogel beads based on N,N,N trimethyl chitosan alginate complexes loaded nano particles. *Chin. Chem. Lett.* 26(9):1129-1132.
- Fei H, Ran Y, Xin C, Jing Y, Yanan C, He Y, Sanming L (2012). Nanostructured lipid carriers (NLC) based topical gel of flurbiprofen: Design, characterization and in vivo evaluation. *Int. J. Pharm.* 439(1-2):15:349-357.
- Harshad V, Abhinesh K, Krutika S (2013). Development of solid lipid nanoparticles based controlled release system for topical delivery of terbinafine hydrochloride. *Eur. J. Pharm. Sci.* 49(2):311-322.
- Isabelle B (2014). Pemulen starch emulsified gel WO2014207715 A2. PCT/IB2014/062662. Dec 31, 2014.
- Lippacher A, Müller RH, Mader K (2001). Preparation of semisolid drug carriers for topical application on solid lipid nano particles. *Int. J. Pharm.* 214:9-12.
- Luigi T, Luciano C, Alberto B, Giuseppe L, Umberto M, Luigi P, Aldo P, Giovanni T, Alberto S (1995). Aceclofenac cream versus piroxicam cream in the treatment of patients with minor traumas and phlogistic affections of soft tissues: a double-blind study. *Curr. Ther. Res.* 56(7):702-712.
- Elgadir Abd, Md.Salim U, Sahena F, Aishah A, Chowdhury AJK, Md. Zaidul Sarker I (2015). Impact of chitosan composites and chitosan nanoparticles composites on various drug delivery systems: A review. *J. Food Drug Anal.* 23(4):619-629.
- Monika S-K, Wolfgang M, Hans-Christian K (2007). Lipid nanoparticles for improved topical application of drugs for skin diseases. *Adv. Drug Del. Rev.* 59(6):427-443.
- Shahin M, Hady SA, Hammad M, Mortada N (2010). Optimized formulation for the topical administration of clotrimazole using Pemulen polymeric emulsifier. *Drug Dev. Ind. Pharm.* 37(5):559-68.
- Sharma R, Walker RB, Pathak K (2011). Evaluation of the kinetics and mechanism of drug release from Econazole nitrate nanosponge loaded carbapol hydrogel. *Int. J. Pharm. Educ. Res.* 45(1):25-31.
- Simovic S, Milic-Askrabic J, Vuleta G, Ibric S, Stupar M (1999). The Influence of Processing Variables on Performance of O/W Emulsion Gels Based on Polymeric Emulsifier (Pemulen (R)TR-2NF). *Int. J. Cosmet. Sci.* 21(2):119-25.



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